

# Silver nanoparticles misting - an innovative method of archaeological object disinfection

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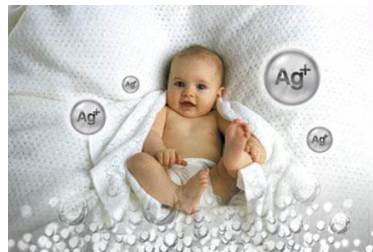


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• Visegrad Fund  
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*Research co-funded by the International Visegrad Fund  
Modern approach for biodeterioration assessment and disinfection of historical book collections*

# Silver nanoparticles application

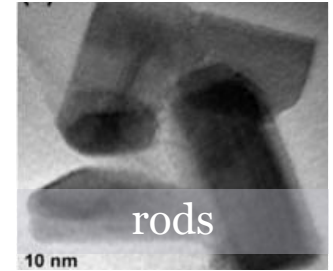
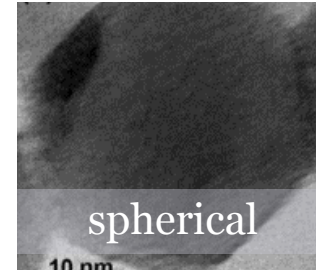
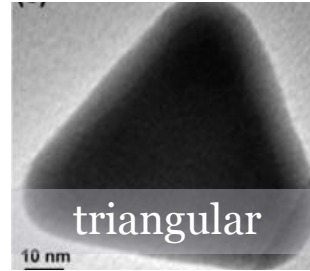
- filters for air purification
- filters for water purification
- stomatology
- dietary supplements
- implants
- textiles
- cosmetics, detergents
- home appliance
- computer hardware
- coatings, grouts, adhesives
- food packaging
- papermaking
- disinfection of historical object



# Silver nanoparticles (AgNPs)

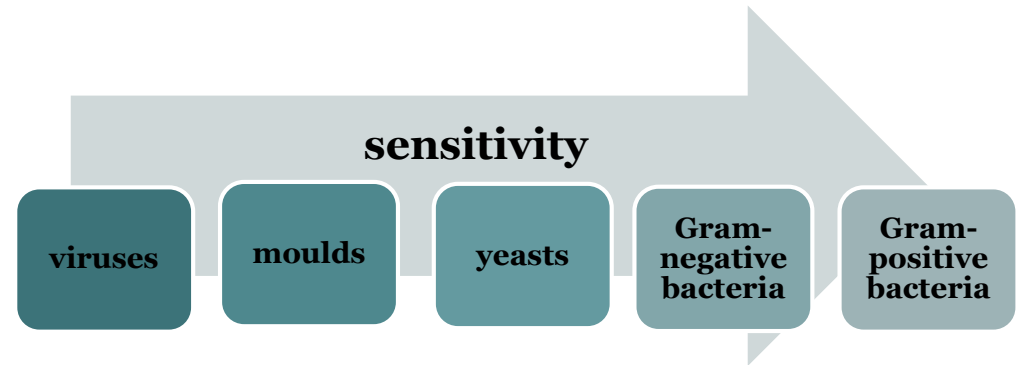
## Synthesis

- chemical
- physical
- biological

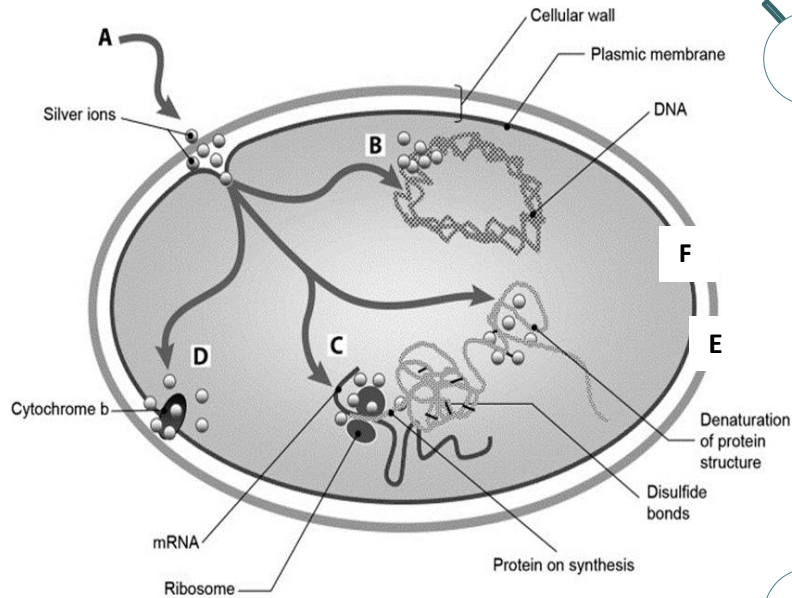


## Effectiveness

- shape
- form
- size
- concentration
- microorganisms



# Mechanism of antibacterial action of AgNPs



**A** disfunction of transport to the cell

**B** inhibition of DNA replication

**C** loss of the biological activity of the amino acid

**D** structural and functional changes in the cell membrane

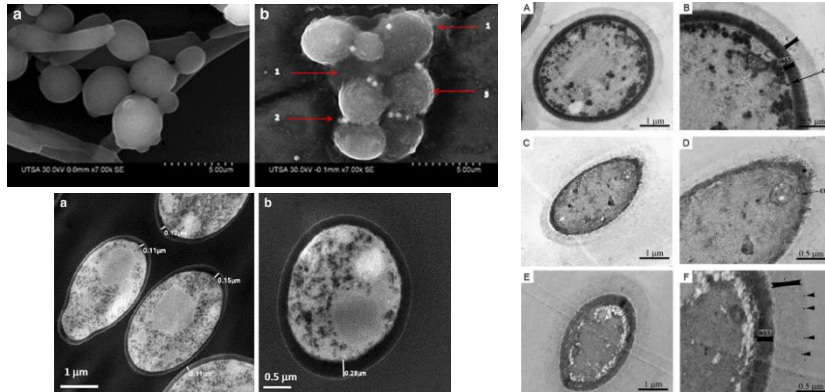
**E** disturbance of the cell membrane electric potential

**F** outflow of protons and certain metabolites through the cytoplasmic membrane

# Mechanism of antifungal action of AgNPs

## Yeasts

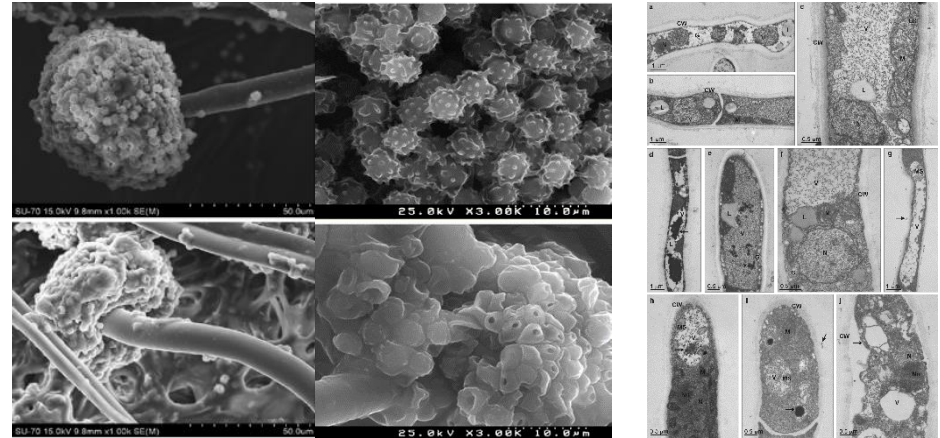
- changes of cell wall functions and structure
- creation of "pits" in the cell membrane
- disturbances of electric potential
- inhibition of budding process



*Candida albicans* and *Cryptococcus neoformans*  
after AgNPs treatment

## Moulds

- inhibition of sporulation process
- changes in cell ultrastructure
- changes in metabolome (acids, enzymes, proteins)



*Aspergillus niger* after AgNPs treatment

# Aim and scope

Optimization of AgNPs misting disinfection and its influence on microorganisms and disinfected materials.

1. Determination of the sensitivity of pure culture collection microorganisms and isolates from historical objects surfaces on the silver nanoparticles preparation.
2. Parameters optimization of the of effective disinfection of technical and historical materials with silver nanoparticles misting.
3. Assessment of the effectiveness and durability of disinfection depending on the microorganism, technical or historical material.
4. Analysis of the disinfection influence on mechanical and optical parameters of technical materials before and after artificial ageing.

# Materials

## Technical:

- paper\* (Sa, Hmp, Sy, CTMP, GW)
- leather (cowhide, dyed)
- textiles (wool, silk, cotton, linen)
- wood (beech, oak, pine)

## Historical:

- canvas (historical painting)
- parchment (conservation materials)
- wood (church floor; 17th c.)
- paper (map; 18th - 19th c.)
- textiles (wool, cotton, sisal; 13th – 15th c.)
- ceramics (6th – 13th c.)



Wool



Ceramics



Cotton

\*Sa - bleached pine kraft pulp; Hmp - bleached hemp kraft pulp; Sy - bleached spruce sulphite pulp; CTMP - chemi-thermomechanical pulp; GW - bleached groundwood from spruce (GW)

# Silver nanoparticles

## AgNPs 1

chemical reduction  
AgNO<sub>3</sub>; reductor:  
sodium citrate;  
stabilizer: PVP

concentration:  
90 ppm

particle size:  
10-80 nm  
(10-15 nm – 60-70%;  
50-80 nm – 30-40%)

Mennica Polska  
S.A.

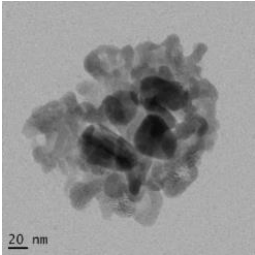
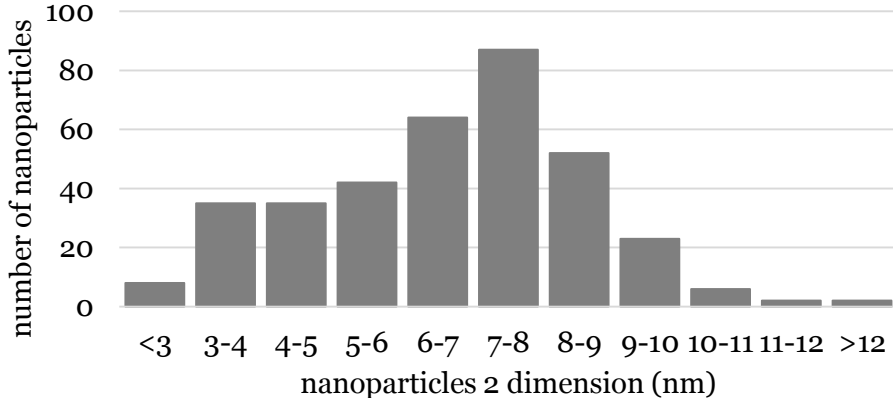
## AgNPs 2

thermal  
decomposition  
of silver  
compounds;  
stabilizer: paraffins

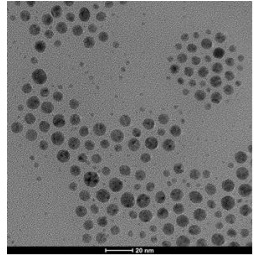
concentration:  
1000 ppm

particle size:  
3-8 nm

Amepox sp. z o.o.



SEM image  
of nanosilver 1



TEM image  
of nanosilver 2



# Microorganisms

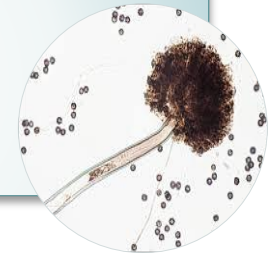
- *Aneurinibacillus aneurinilyticus*
- *Brevibacillus laterosporus*
- *Bacillus subtilis*
- *Bacillus megaterium*
- *Bacillus pumilus*
- *Bacillus licheniformis*
- *Sphingomonas paucimobilis*
- *Micrococcus* sp.
- *Micrococcus flavus*
- *Pseudomonas aeruginosa*
- *Staphylococcus aureus*
- *Staphylococcus lentus*
- *Staphylococcus xylosus*
- *Nocardia* sp.
- *Escherichia coli* (ATCC 10536)
- *Staphylococcus aureus* (ATCC 6538)
- *Bacillus subtilis* (NCAIM 01644)

## BACTERIA



- *Alternaria alternata*
- *Aspergillus versicolor*
- *Aspergillus niger*
- *Cladosporium cladosporioides*
- *Cladosporium herbarum*
- *Cladosporium macrocarpum*
- *Mucor racemosus*
- *Penicillium digitatum*
- *Penicillium carneum*
- *Penicillium crustosum*
- *Penicillium radicola*
- *Rhizopus nigricans*
- *Candida sphaerica*
- *Rhodotorula* sp.
- *Rhodotorula mucilaginosa*
- *Aspergillus niger* (ATCC 16404)
- *Penicillium chrysogenum* (ŁOCK 0531)
- *Candida albicans* (ATCC 10231)

## FUNGI



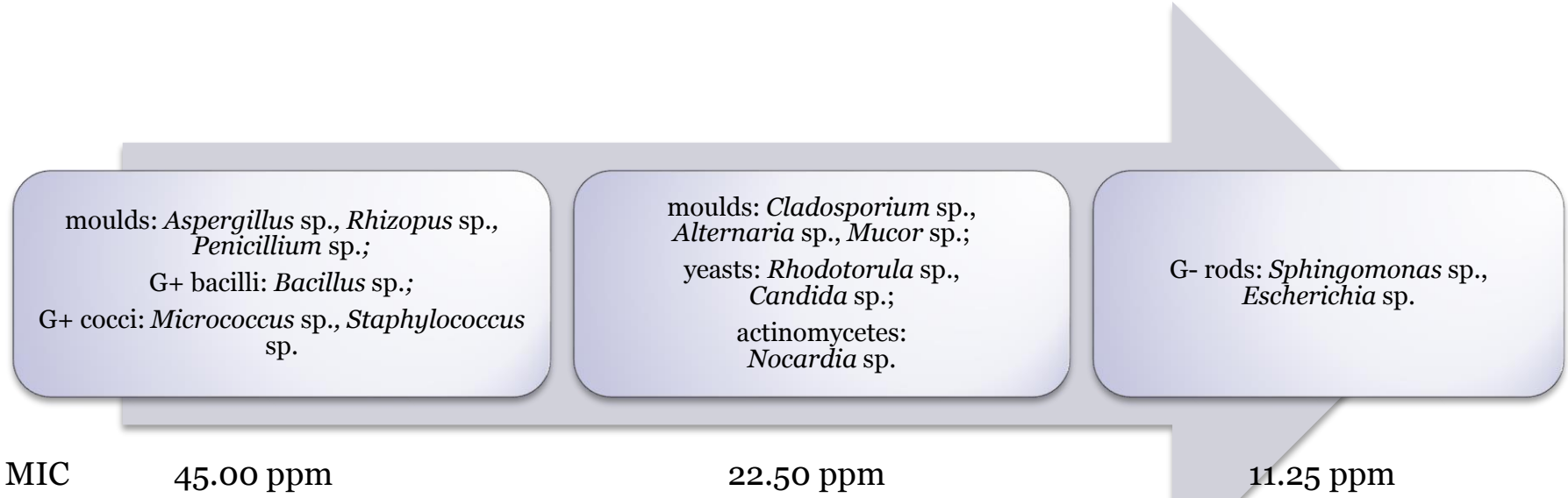
# Methods

Analysis	Method / Standard
<b>Silver nanoparticles</b>	
Characterization of preparation	SEM; TEM
Determination of AgNPs content	FAAS; LA-ICP-TOF-MS
<b>Microorganisms</b>	
Determination of microbial sensitivity (MIC and MBC)	ASTM E2149 – 01
Choosing the conditions for disinfection Evaluation of the effectiveness and durability of the disinfection technical materials and historical objects	AATCC Test Method 100-2012
<b>Materials</b>	
Assessment of mechanical parameters (breaking, elongation, tear, compression)	ISO 13934-1:1999 (textiles); ISO 3376:2005 (leather); ISO 20187:1993 (paper); ISO 1924-1:1998 (paper); BS EN 21974:2002 (paper); BS EN 384:2004 (wood)
Estimation of colour parameters	Metoda Tappi T 524 om-94 (paper); ISO 2470:1999 (paper); ISO 105-J01:2002 (leather, textiles); Metoda SCI (wood)

# Microbial sensitivity to AgNPs

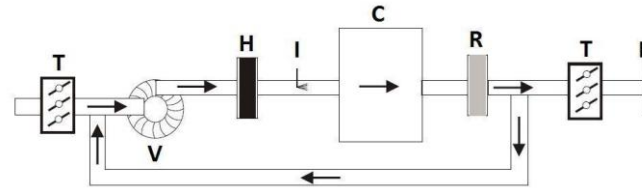
Microorganisms	Origin	MIC (ppm)	MBC (ppm)
<i>Staphylococcus aureus</i>	ATCC	11.25	22.50
	Museum 1	22.50	45.00
<i>Bacillus subtilis</i>	ATCC	22.50	> 45.00
	Archives	22.50	> 45.00
	Museum 2	45.00	> 45.00
<i>Escherichia coli</i>	ATCC	11.25	22.50
<i>Nocardia</i> sp.	Library	11.25	22.50
<i>Aspergillus niger</i>	ATCC	22.50	45.00
	Museum 2	45.00	45.00
<i>Candida albicans</i>	ATCC	11.25	22.50
<i>Candida sphaerica</i>	Library	22.50	22.50

# Microbial sensitivity to AgNPs



# AgNPs misting chamber

**Capacity: 1.73 m<sup>3</sup> → 36-42 books A4**



T – throttling valve; V – ventilator; H – heater;  
I – silver colloid injection; C – chamber; R – radiator; F – filter

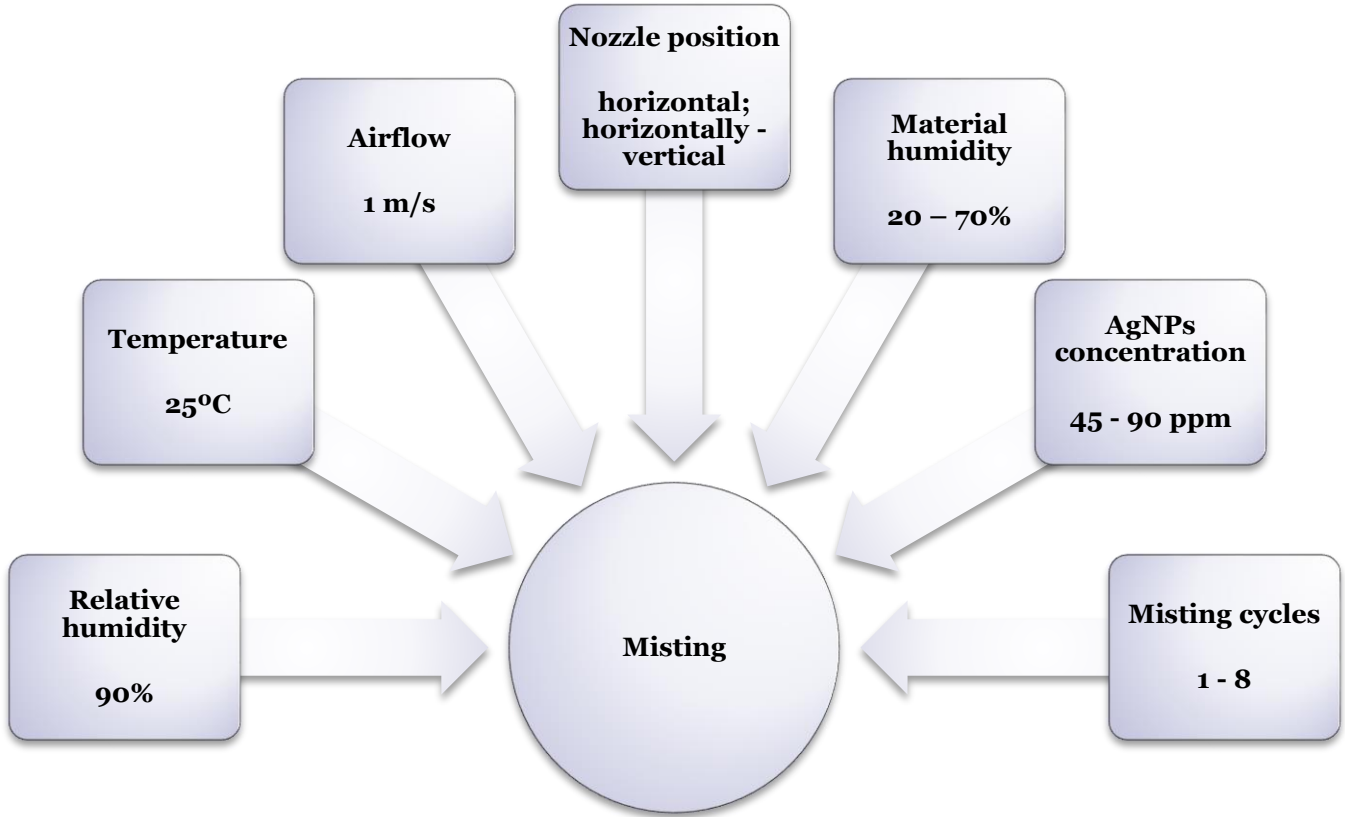
Wasiak R., Laskowski Z., Czyżyk J. The microbiological protection method of archive and museum objects and installation for the microbiological protection of archive and museum objects. Patent PL399507, Poland 2012



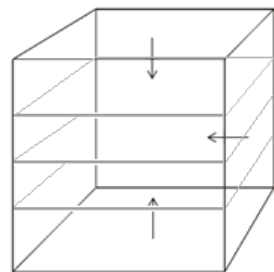
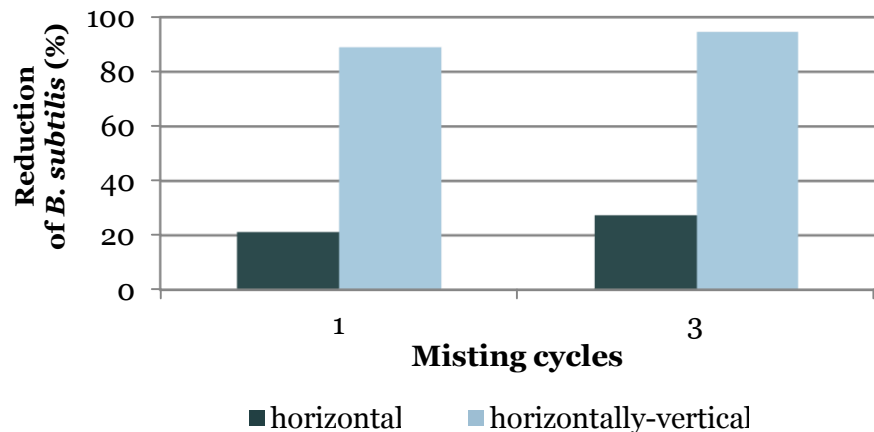
INSTAL WARSZAWA S.A.

Siennicka 29, 04-394 Warsaw, Poland  
[www.instalwaw.com.pl](http://www.instalwaw.com.pl)

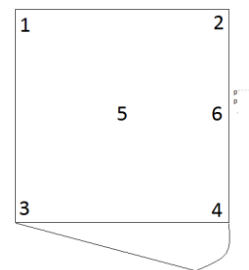
# Choosing the conditions for AgNPs misting



# Nozzle position and AgNPs distribution



Nozzle position  
(horizontally – vertical)

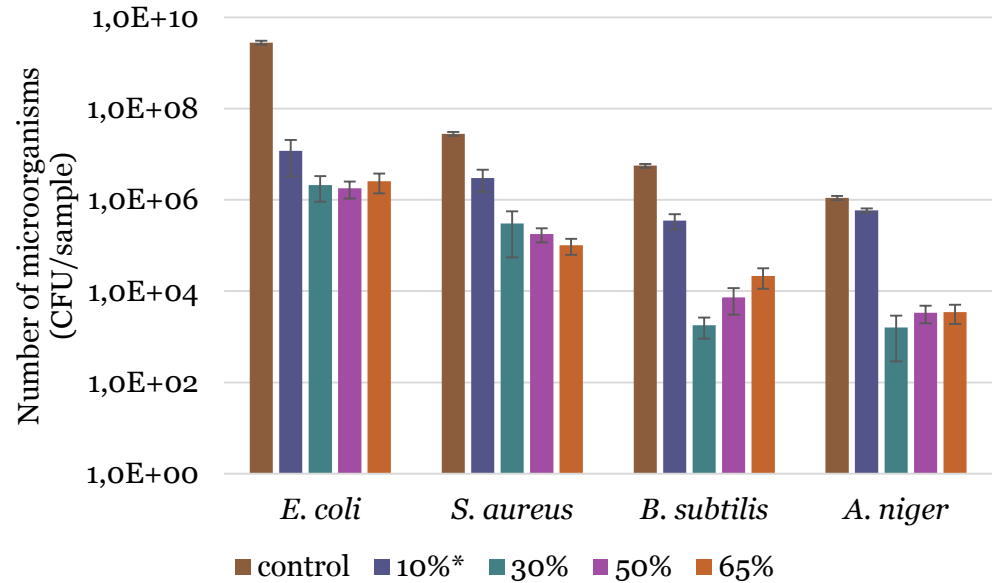
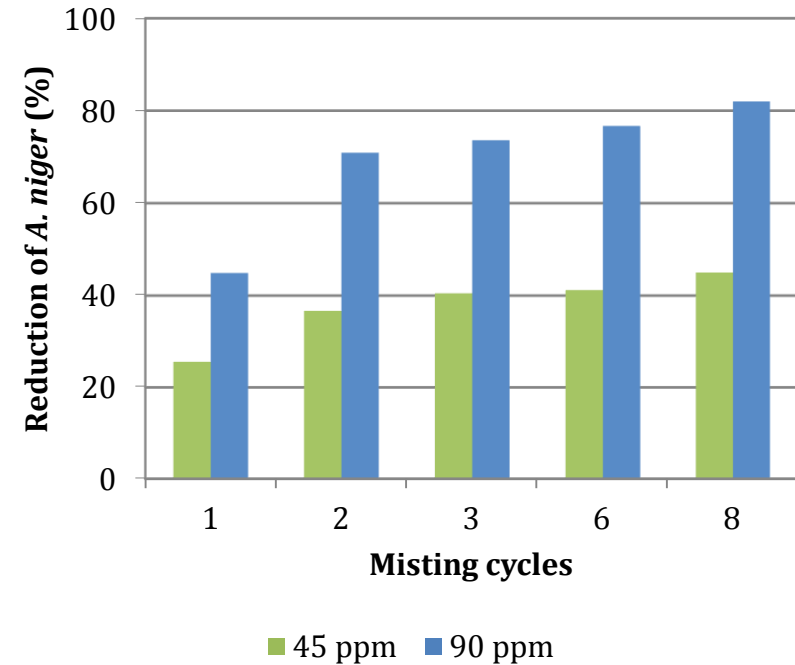


Sample points  
distribution

Sampling point	AgNPs amount (ppm/g)*
1	2.8
2	2.2
3	3.3
4	2.7
5	2.9
6	2.3

\* FAAS – Flame Atomic Absorbance Spectrometry

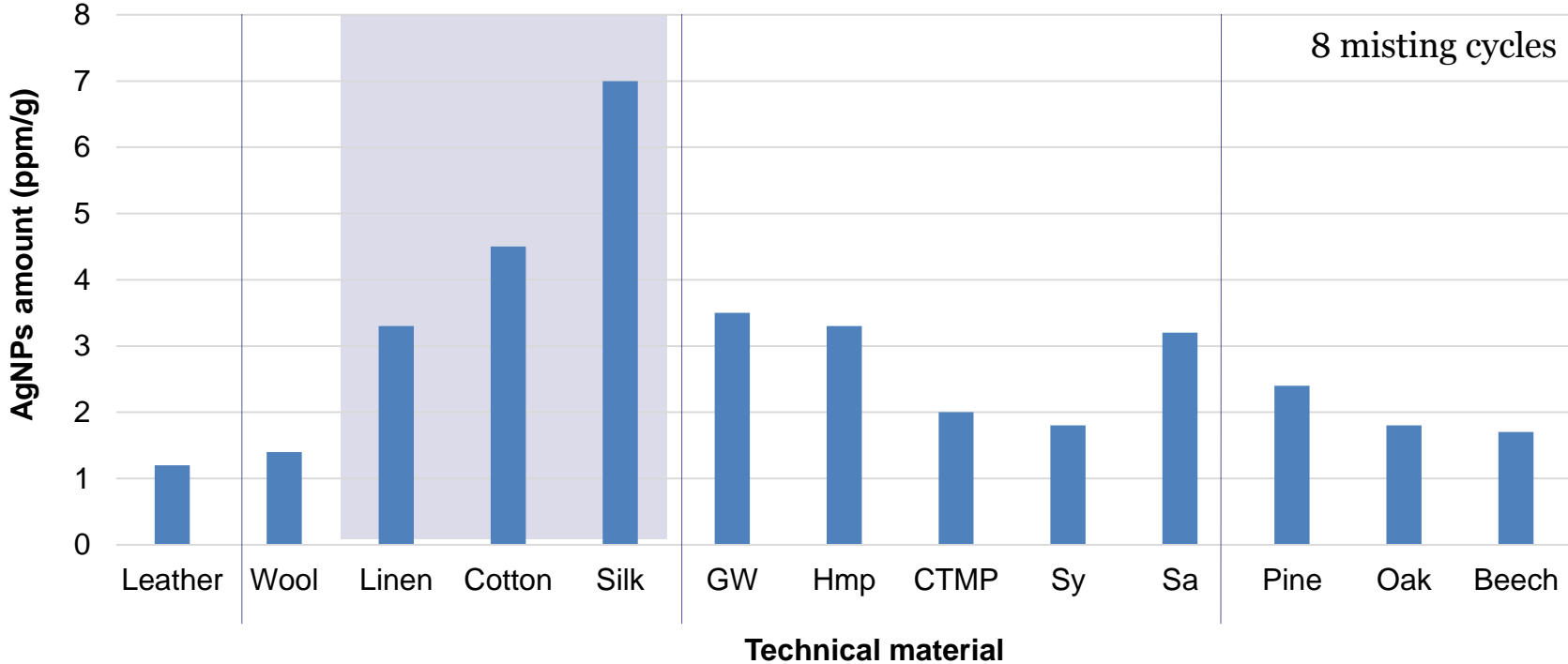
# Misting cycles, AgNPs concentration, material humidity



10% - dry cotton



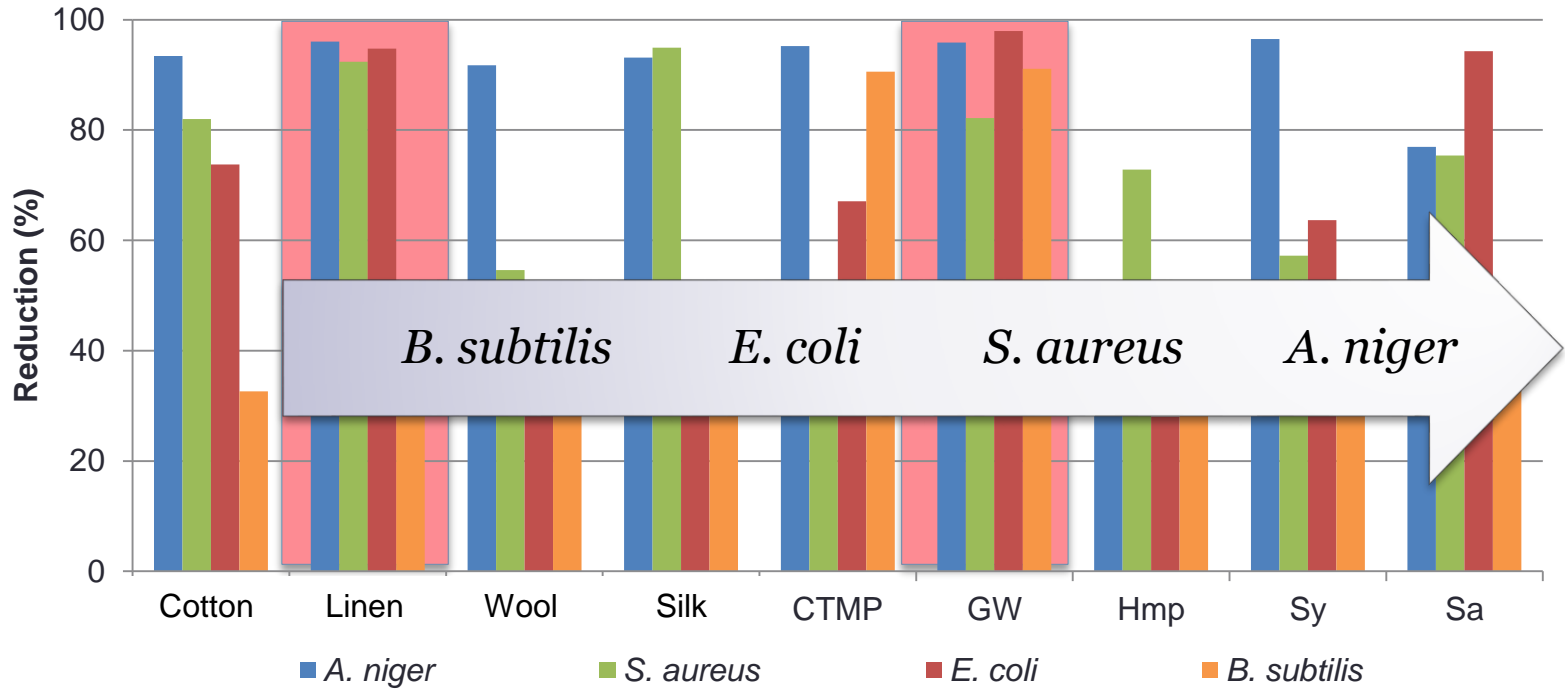
# Silver nanoparticles amount by FAAS



FAAS – Flame Atomic Absorbance Spectrometry; Sa - bleached pine kraft pulp; Hmp - bleached hemp kraft pulp; Sy - bleached spruce sulphite pulp; CTMP - chemi-thermomechanical pulp; GW - bleached groundwood from spruce (GW)

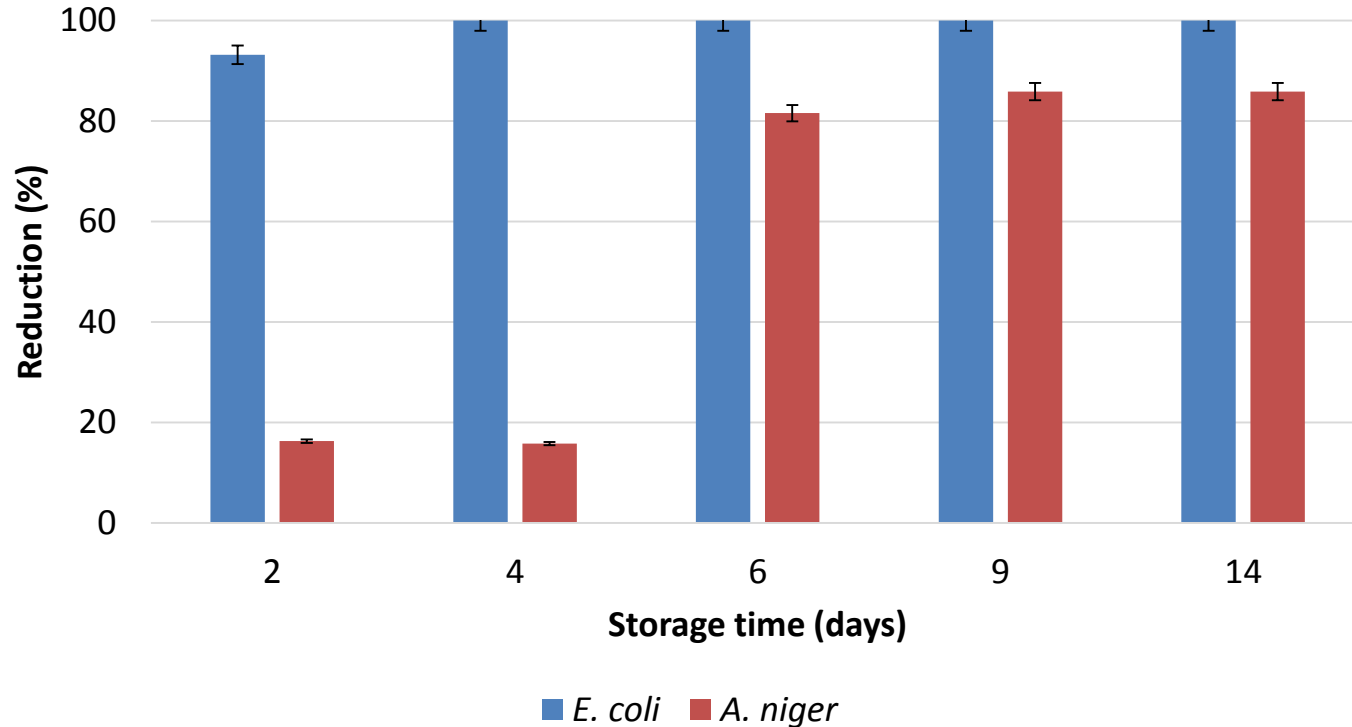
Gutarowska et al., 2014

# Disinfection effectiveness on technical materials



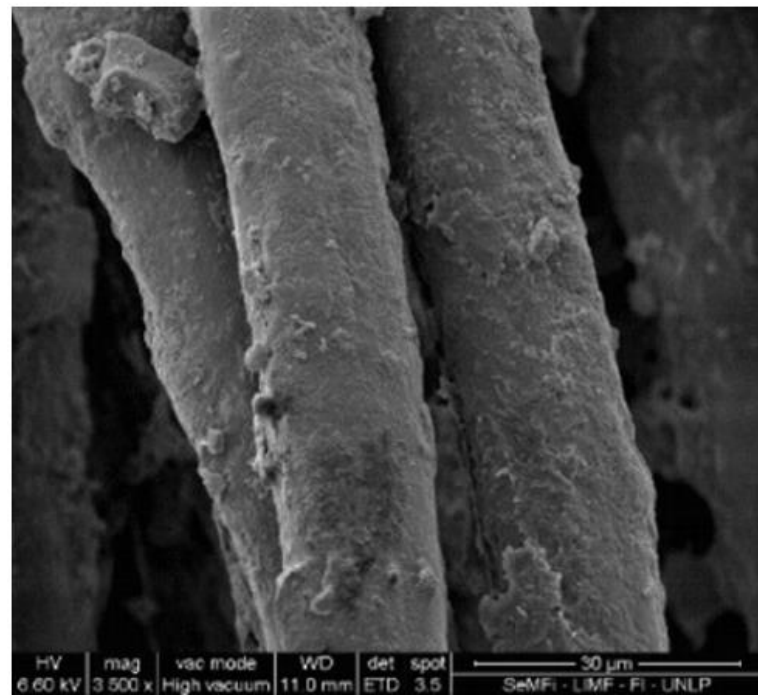
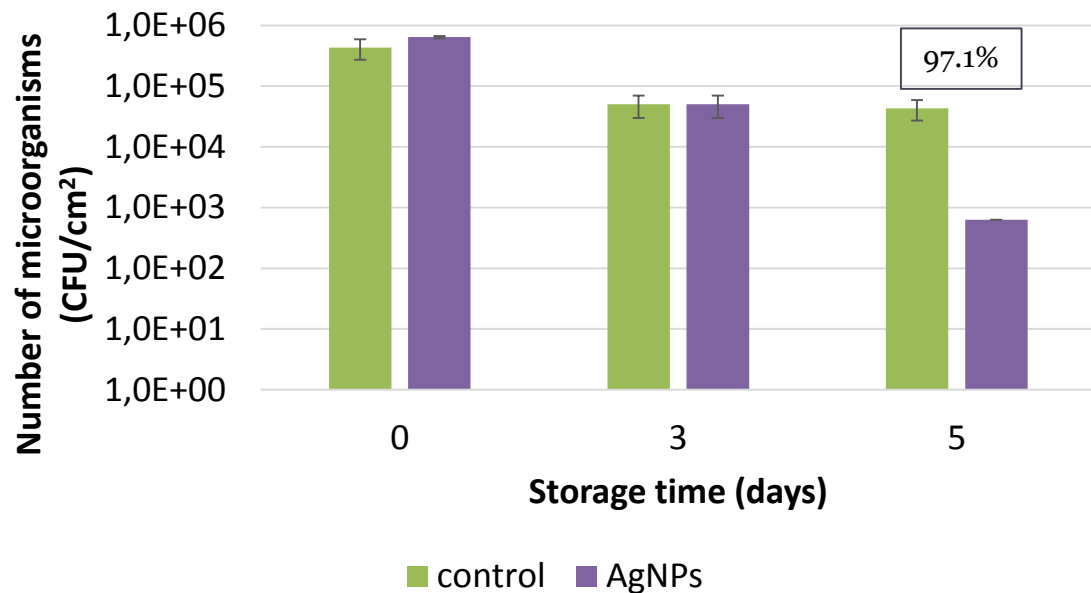
\*Sa - bleached pine kraft pulp; Hmp - bleached hemp kraft pulp; Sy - bleached spruce sulphite pulp; CTMP - chemi-thermomechanical pulp; GW - bleached groundwood from spruce (GW)

# AgNPs misting durability



Storage conditions: 28°C, RH 80%

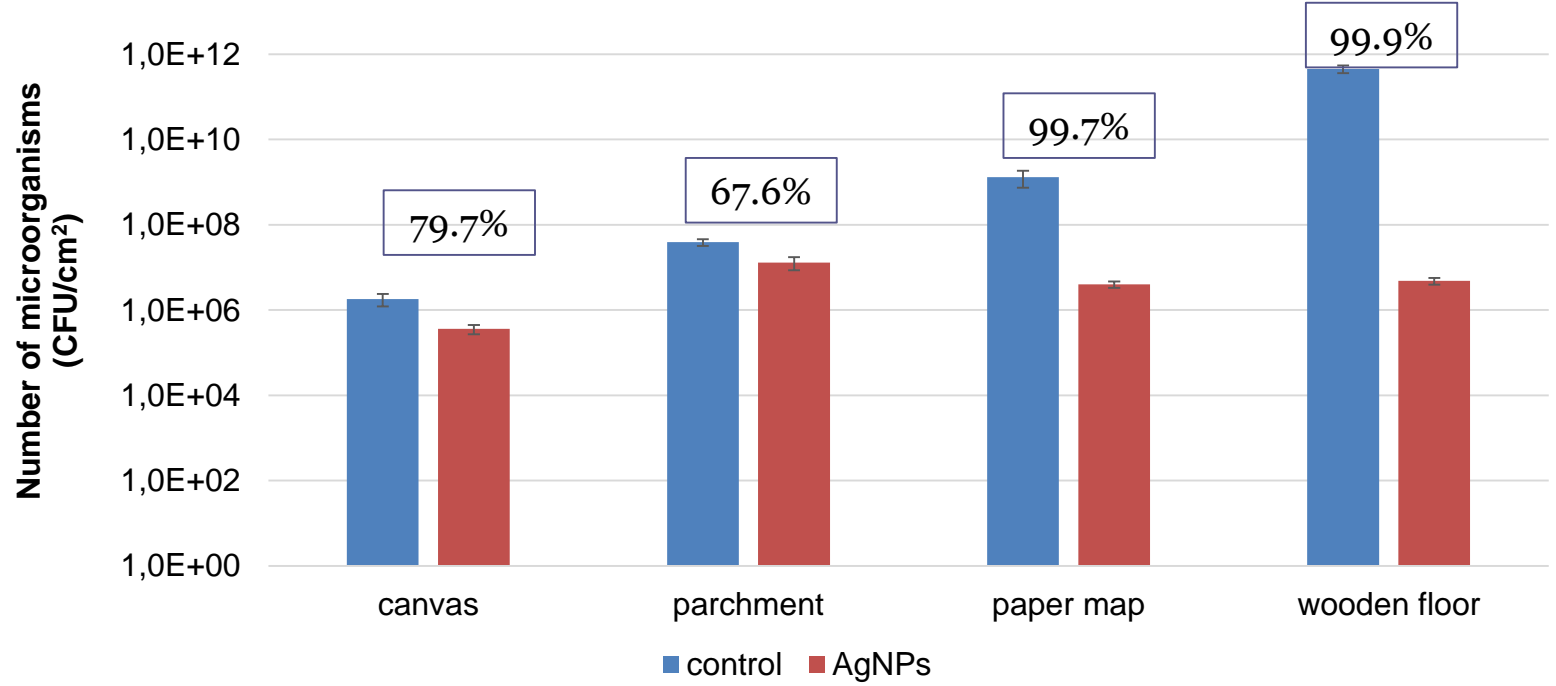
# Inhibition of *Pseudomonas aeruginosa* growth



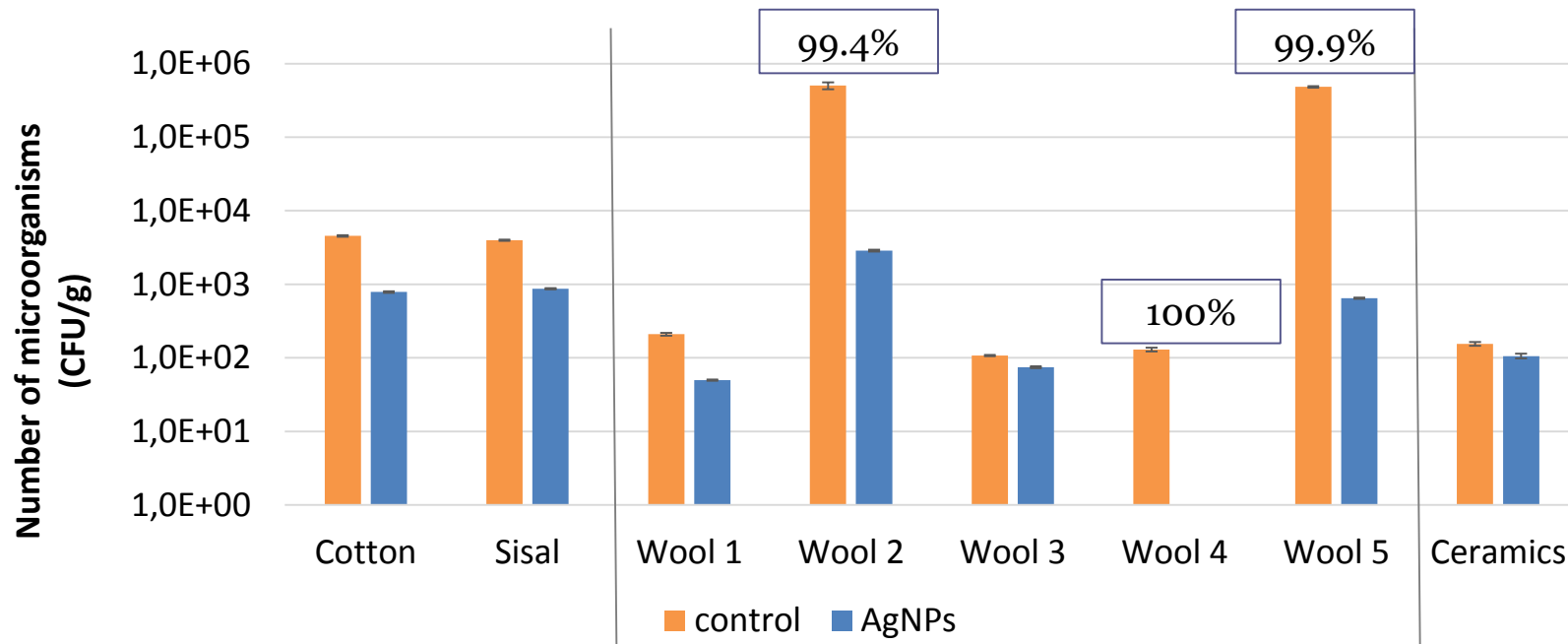
*Pseudomonas aeruginosa* on historical textile

Storage conditions: 28°C, RH 80%

# Disinfection effectiveness on historical materials



# Disinfection effectiveness on archaeological textiles



# Mechanical parameters of paper

Paper	Control		AgNPs		Aged control		Aged AgNPs	
	Elongation (%)	Tear index (mN×m <sup>2</sup> /g)	Elongation (%)	Tear index (mN×m <sup>2</sup> /g)	Elongation (%)	Tear index (mN×m <sup>2</sup> /g)	Elongation (%)	Tear index (mN×m <sup>2</sup> /g)
<b>GW</b>	2.4±0.2	8.1±0.3	2.8±0.2	8.1±0.5	2.2±0.1	7.5±0.2	2.1±0.2	7.2±0.8
<b>Sy</b>	2.0±0.1	9.3±0.8	2.4±0.2	8.6±0.4	1.9±2.2	7.6±0.4	1.8±0.2	7.2±0.3
<b>Sa</b>	3.0±0.2	13.5±0.9	3.1±0.2	13.9±0.4	2.6±0.1	12.9±0.3	2.6±0.1	12.8±0.4

Sa - bleached pine kraft pulp; Sy - bleached spruce sulphite pulp; GW - bleached groundwood from spruce (GW)

# Optical parameters of paper

Paper	Control	AgNPs		Aged control		Aged AgNPs	
	R457	R457	$\Delta E$	R457	$\Delta E$	R457	$\Delta E$
<b>GW</b>	82.9±0.1	82.1±0.1	0.41	78.6±0.5	1.02	78.5±0.1	0.94
<b>Sy</b>	89.8±0.5	89.4±0.3	1.06	85.4±0.6	0.37	85.5±0.7	1.27
<b>Sa</b>	90.5±0.3	89.6±0.1	0.91	87.7±0.2	0.57	87.7±0.1	0.61

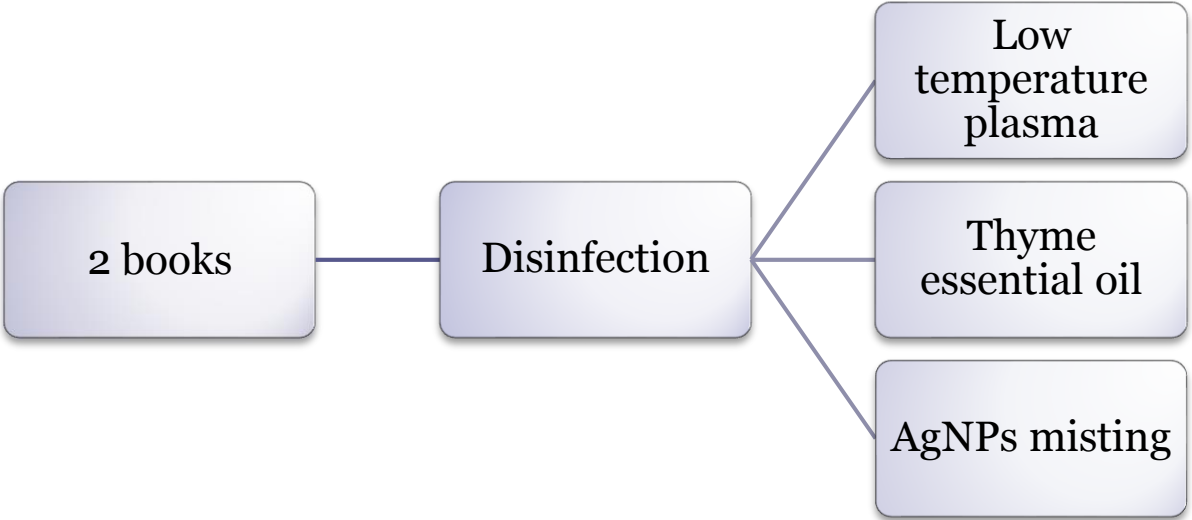
R457 – ISO Brightness; Sa - bleached pine kraft pulp; Sy - bleached spruce sulphite pulp; GW - bleached groundwood from spruce

- 0 <  $\Delta E$  < 1 - observer does not notice the difference
- 1 <  $\Delta E$  < 2 - only experienced observer can notice the difference
- 2 <  $\Delta E$  < 3.5 - unexperienced observer also notices the difference
- 3.5 <  $\Delta E$  < 5 - clear difference in colour is noticed
- 5 <  $\Delta E$  - observer notices two different colours

$$\Delta E = \sqrt{(\Delta L^*)^2 + (\Delta a^*)^2 + (\Delta b^*)^2}$$



# Disinfection of historical book collections



## AgNPs misting

Nanosilver concentration  
90 ppm

Misting cycles  
8

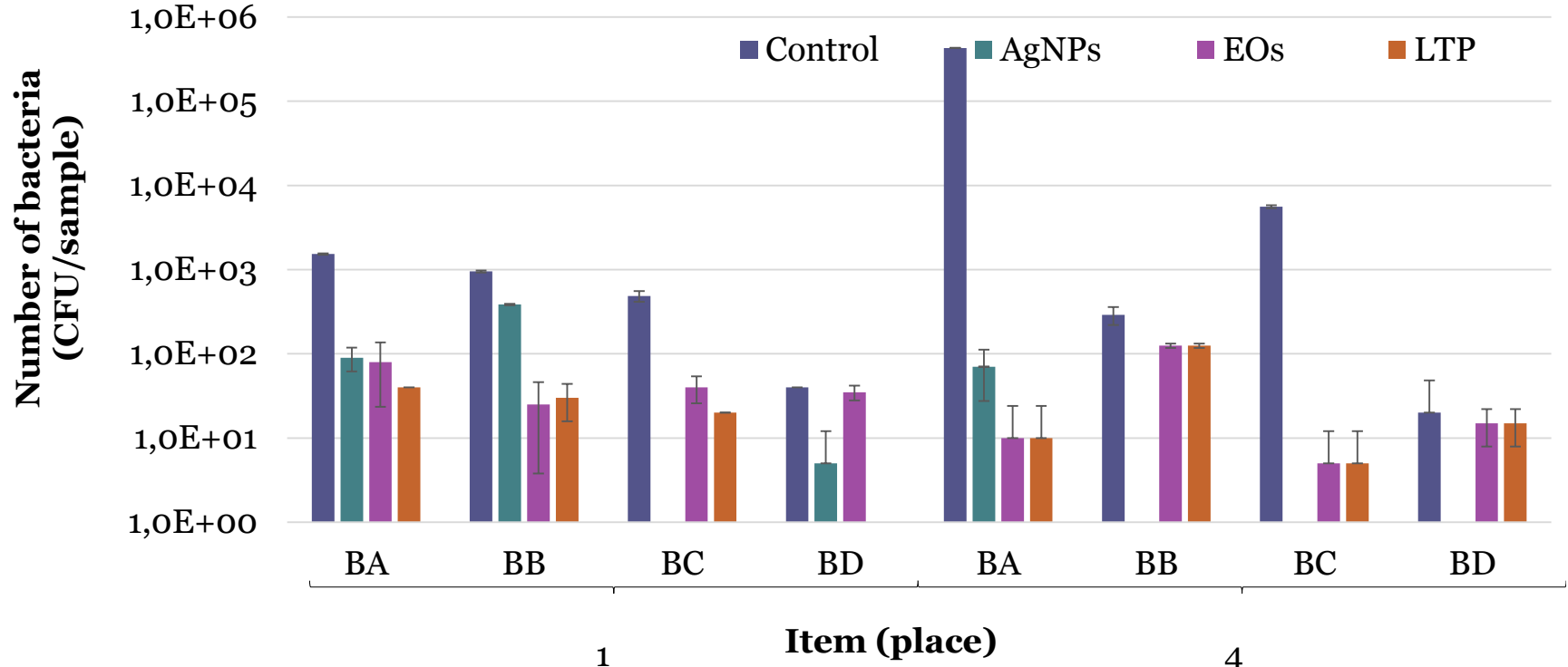
Airflow  
1 m/s

Temperature  
25°C

RH  
90%

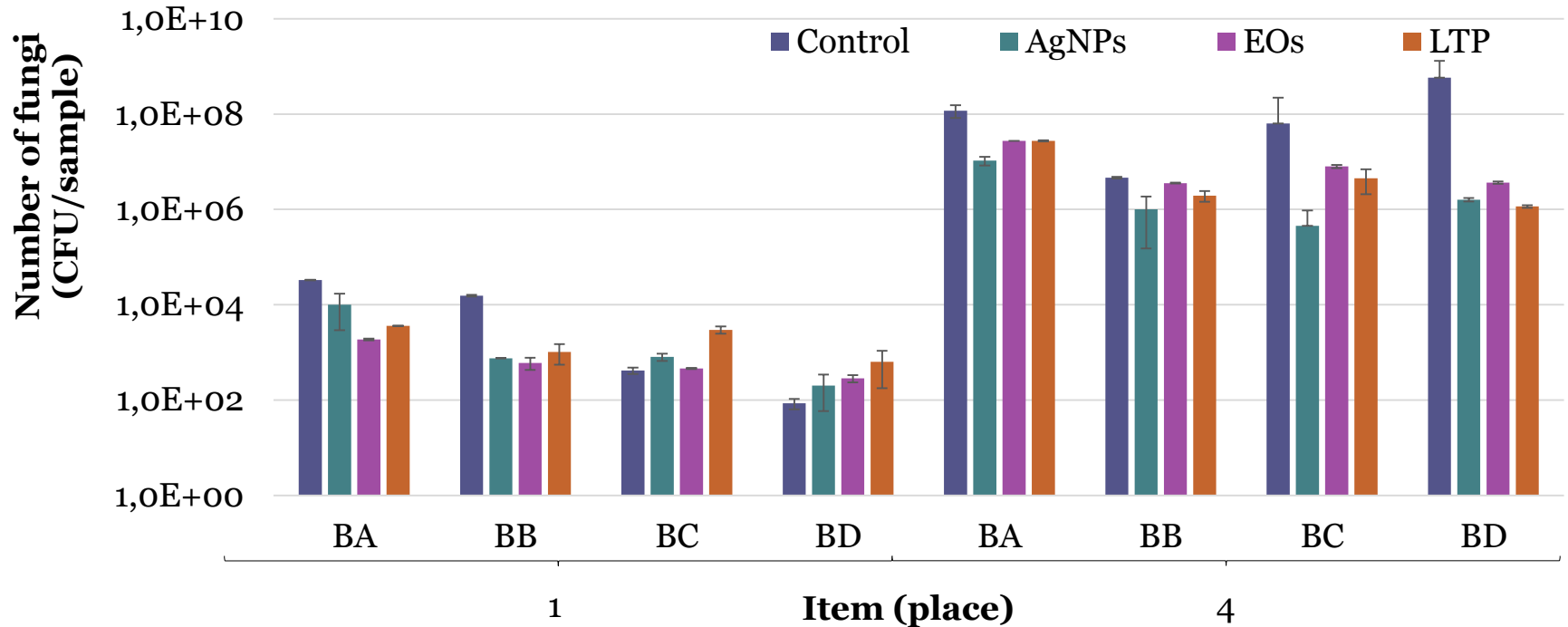
Duration  
520 min

# Disinfection effectiveness: culture-dependent analysis



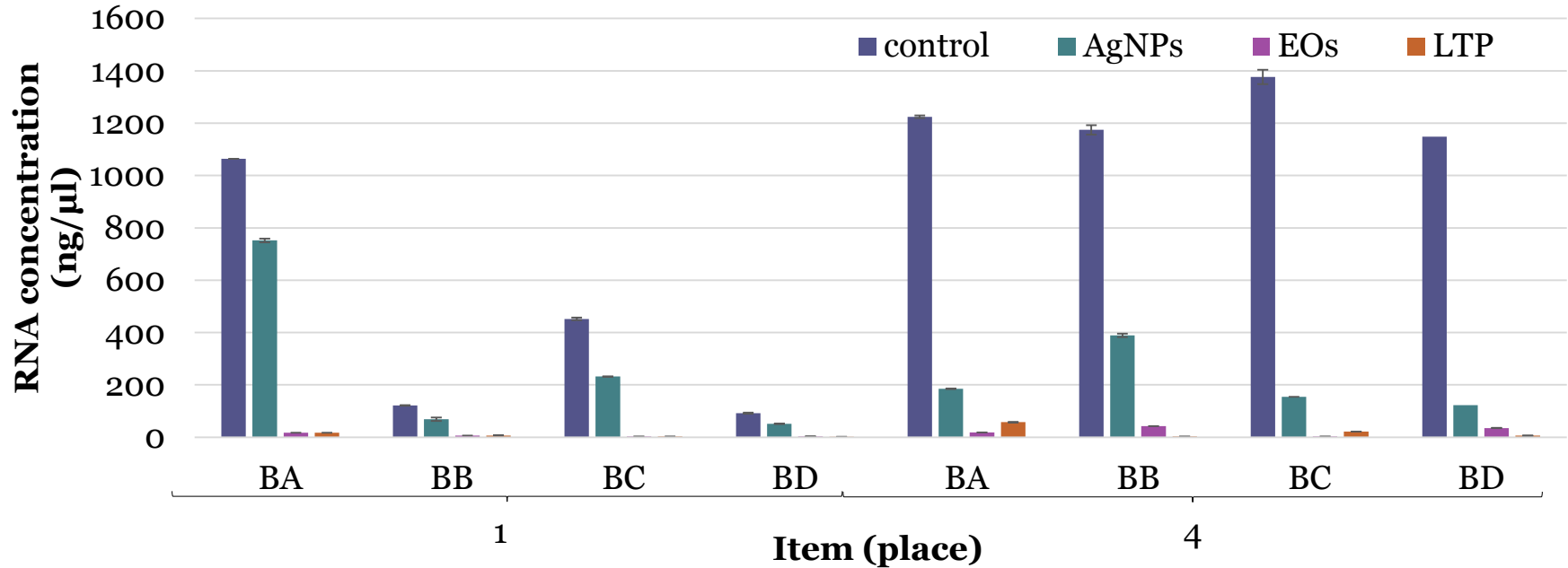
BA – front cover; BB - front free endpaper; BC – fore edge; BD – page inside the book; sample = 25 cm<sup>2</sup>

# Disinfection effectiveness: culture-dependent analysis



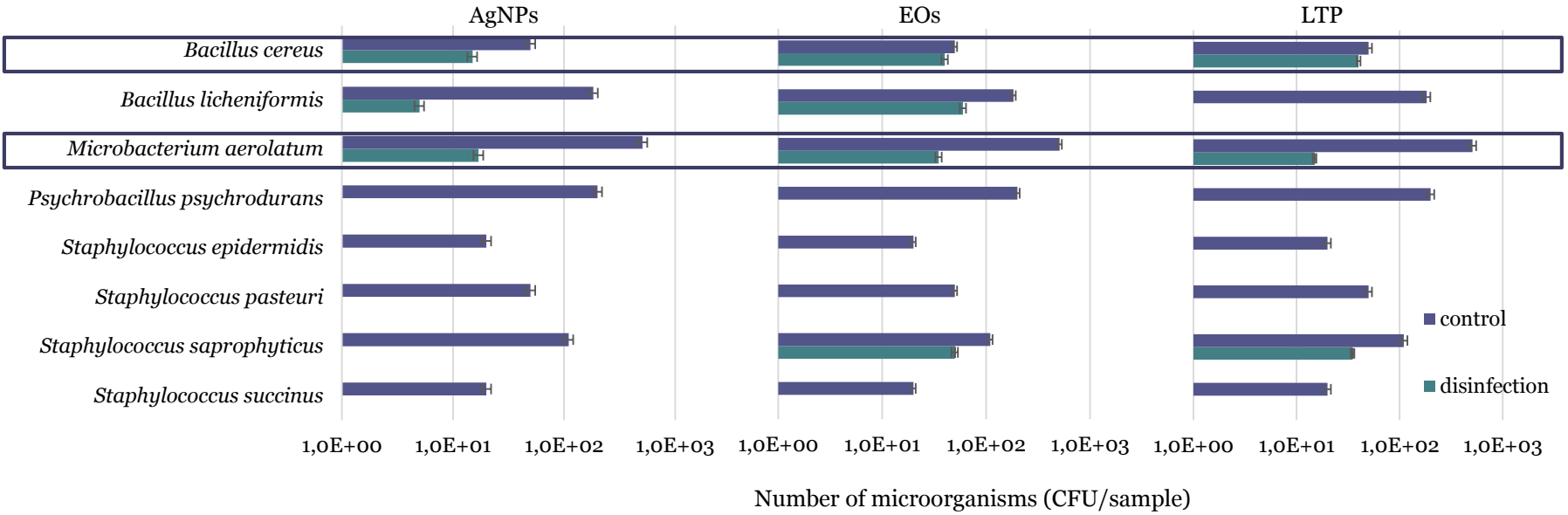
BA – front cover; BB - front free endpaper; BC – fore edge; BD – page inside the book; sample = 25 cm<sup>2</sup>

# Disinfection effectiveness: culture-independent analysis

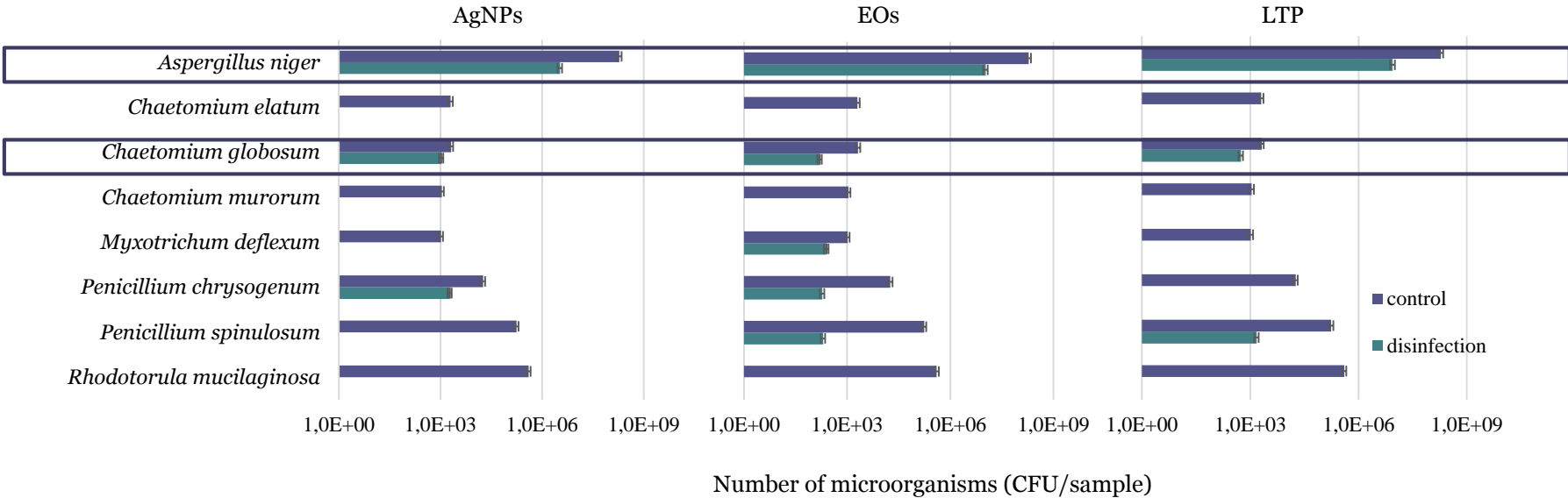


BA – front cover; BB - front free endpaper; BC – fore edge; BD – page inside the book; sample = 25 cm<sup>2</sup>

# Bacterial sensitivity



# Fungal sensitivity



# Conclusions

1. Silver nanoparticles misting, low temperature plasma and thyme essential oil microatmosphere were efficient methods of growth inhibition of bacteria and fungi.
2. The disinfection effectiveness depended on the inhabiting microbiota, disinfected book and the sampling place.
3. The reduction of microorganism number equalled on Book no. 1 to: AgNPs and LTP (0–100%), EOs (0–97 %); and on Book no. 4 to: AgNPs 78–100%, LTP (25–100%) EOs 23–100%).
4. RNA concentration is a good marker to assess the disinfection effectiveness.
5. *Aspergillus niger*, *Chaetomium globosum*, *Bacillus cereus* and *Microbacterium aerolatum* were insensitive to all disinfection methods.